Macroinvertebrates and Microbes (Archaea, Bacteria) Offer Complementary Insights into Mine-Pit Lake Ecology

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Abstract

The broad objective of this research was to determine the environmental drivers of macroinvertebrate and microbial assemblages in acidic pit lakes. This is important because pit lake ecosystem development is influenced by prevailing environmental characteristics. Three lakes (Stockton, Kepwari, WO5H) within a larger pit-lake district in Collie, Western Australia were surveyed for spatial variability of benthic macroinvertebrate and microbe (Archaea, Bacteria) assemblage composition as well as potential environmental drivers (riparian condition, aquatic habitat, sediments, and aquatic chemistry) of assemblages. With the exception of sediment chemistry, biophysical variables were significantly different across lakes and reflected riparian condition and groundwater chemistry. Microbial assemblages in pit lakes were significantly different across lakes and correlated with water chemistry, particularly metals in Lake WO5H. However, the most abundant microbes were not readily identified beyond class, making it difficult to speculate on their ecological function. Macroinvertebrate assemblage composition and species richness were also significantly different across all lakes, and in Lake WO5H (a lake with low pH and high metal concentrations), taxa were correlated with benthic organic matter as well as water chemistry. Results indicated that despite poor water quality, input of nutrients from terrestrial leaf litter can support or augment pit lake ecosystems. This is a demonstration of the concept that connection of pit lakes to catchments can positively affect aquatic ecosystems, which can inform management actions for remediation.

Keywords Collie · Kepwari · Pit lake · Water quality · Sediment chemistry · Riparian · Closure · Monitoring

Introduction

Large-scale mining permanently changes landscapes. Mountaintop removal, construction of new landforms, and the creation of thousands of open pits occurs across every inhabited continent on earth (Blanchette and Lund 2016). Pit lakes form when these open pits fill with ground, surface and rain water, and by virtue of their size and water toxicity, can have substantial environmental impact (Lund and

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Blanchette 2018; Miller et al. 1996; Younger and Wolkersdorfer 2004). Historically, pit lake rehabilitation has been considered relatively unimportant, with lakes locked away from public access, either mandated for 'perpetual treatment' or simply abandoned with minimal (if any) safety measures (Blanchette and Lund 2016). There is growing recognition that successful mine closure is a stakeholder-driven process (Bainton and Holcombe 2018), and communities are increasingly unwilling to tolerate abandoned pit lakes on their doorstep (see Kean 2009; Woodbury 1998).

By virtue of their location, size, and geology, pit lakes can be considered to inhabit a point along a 'sliding scale' of interacting factors that increases the complexity of rehabilitation (Blanchette and Lund 2016). For example, a large lake with highly toxic water and prone to frequent tropical cyclones may lay at the more 'difficult' end of the scale. However, not all pit lakes are alike. Shallow (1–5 m) coal-strip lakes with large amounts of terrestrial leaf litter (Campbell and Lind 1969; Coe and Schmelz 1972) are now recreational fisheries in the US Midwest. The well-forested

pit lake district of Lusatia, Germany has been developed for public recreational activities such as boating, upmarket hotels, spas, and restaurants, and is projected to contribute at least €10 M to the local economy (Lienhoop and Messner 2009). If these new lakes are to be community assets, they must be properly monitored and understood—beyond simple water quality measures—because community members expect water bodies to be safe, accessible, and biodiverse (sensu Naiman 2013).

Aquatic macroinvertebrates are routinely used to assess the condition of bioindicators of freshwater ecosystems. The ecology of aquatic macroinvertebrates is comparatively well-known, with an extensive body of literature to assist monitoring programs. Advances in technology can enhance monitoring programs by using DNA sequencing to identify macroinvertebrates (Carew et al. 2013) or measure gene expression to determine environmental stress (Chou et al. 2018). Increasingly, microbial assemblages (16S) are being used either alone or in conjunction with macroinvertebrates to characterise freshwater ecosystem condition and response to anthropogenic pressures (e.g. Horton et al. 2019; Simonin et al. 2019; Vander Vorste et al. 2019).

Relative to other inland waterbodies, the ecology of pit lakes is understudied and underrepresented in the primary literature (Blanchette and Lund 2016). As products of the mining industry, pit lake microbial ecology is readily viewed in terms of microbially-mediated biogeochemistry, and understanding these reactions can be used to improve water quality modelling (Wielinga 2009). Past studies used culture-dependent methods or cell counts to characterise communities, inadvertently missing much of the taxonomic diversity. More recent pit lake studies have adopted culture-independent 16S sequencing, which has expanded knowledge of mine pit lake microbial biodiversity (see González-Toril et al. 2015; Kampe et al. 2010; Lucheta et al. 2013), but few studies have compared the drivers of microbes and macroinvertebrates in pit lakes, or applied this knowledge to closure planning.

The broad objective of this research was to determine the drivers of macroinvertebrate and microbial assemblages in acidic pit lakes. This is important because pit lake ecosystem development is influenced by prevailing environmental characteristics (e.g. riparian condition, aquatic chemistry), which inform remediation, monitoring, and closure. Microbes are key to pit lake ecosystems; DNA sequencing technology has become more routine, but collection and analysis is still highly variable and there are currently no standard methods. This work will compare the environmental drivers of microbial assemblages (a relatively new bioindicator) with aquatic macroinvertebrates (an established bioindicator).

The specific aims of the research were to: (1) characterise the biophysical variables of three co-occurring mine pit lakes in Collie, Western Australia, (2) collect macroinvertebrates and microbes (Archaea, Bacteria) from the pit lakes, (3) determine if assemblages were significantly different among lakes, (4) seek biophysical variables that correlated with assemblages, and (5) ascertain if, in relation to macroinvertebrates, microbial assemblages provide alternative or complementary understanding of pit lake ecosystems.

Methods

Site Description

The three study lakes (Kepwari, Stockton, and WO5H) are former coal pits located in the Collie basin, Western Australia (Fig. 1). The coalfield itself is approximately 224 km^2 with Permian sedimentary succession up to 1400 m thick—the top 900 m containing coal seams between 0.5 and 13 m thick (Millar et al. 2011). Mining began by hand in 1892; the first open cut operation started in 1943 (Millar et al. 2011), eventually creating a 'district' of 10 pit lakes (Fig. 1) variably hydrologically connected to the wider catchment by seasonal surface and ground water flows. Collie has a Mediterranean climate with dry summers (14–30 °C) and wet, cold winters (4–16 °C) (mean rainfall for years 2002–2019 was 711 mm; May–Sept was 539 mm; Australian Bureau of Meteorology 2019).

Lake Stockton (Fig. 2) was opened to underground mining in 1927 and open-cut operations began in 1943, with both types of mining active until it was abandoned in 1960 (Stedman 1988). After abandonment, the pit filled naturally with groundwater, creating a lake 25–47 m deep that is currently used for public recreation, particularly water skiing. Lake Stockton has occasionally received dewatering water from Griffin Coal operations, and the lake water periodically discharges into a creek that meets the Collie River South (see Lund and Blanchette 2018).

Pit WO5H (Fig. 2) was mined from 1986 to 1997 and then rapidly filled with surface water until 2004 (McCullough et al. 2009). An acidic (pH 4.9), saline stream flows into Lake WO5H due to seasonal rainfall events, and the lake can discharge, with lake water joining flows from Lake Stockton before running into the Collie River South (McCullough et al. 2009). Discharge from WO5H is prevented in most years by pumping substantial volumes of lake water for use in power station cooling. Efforts were made to plant trees and shrubs below the final water line before filling to add carbon to the highly oligotrophic lake (Lund et al. 2014).

The Collie River South was diverted around the WO5B pit to allow mining. Mining in the WO5B pit ceased in 1996, and it was rapidly filled from 1999 to 2004 from the Collie River South to form Lake Kepwari (Fig. 2), which is 65 m deep and covers an area of 1.03 km^2 (McCullough et al. 2009). The overburden dumps have been extensively
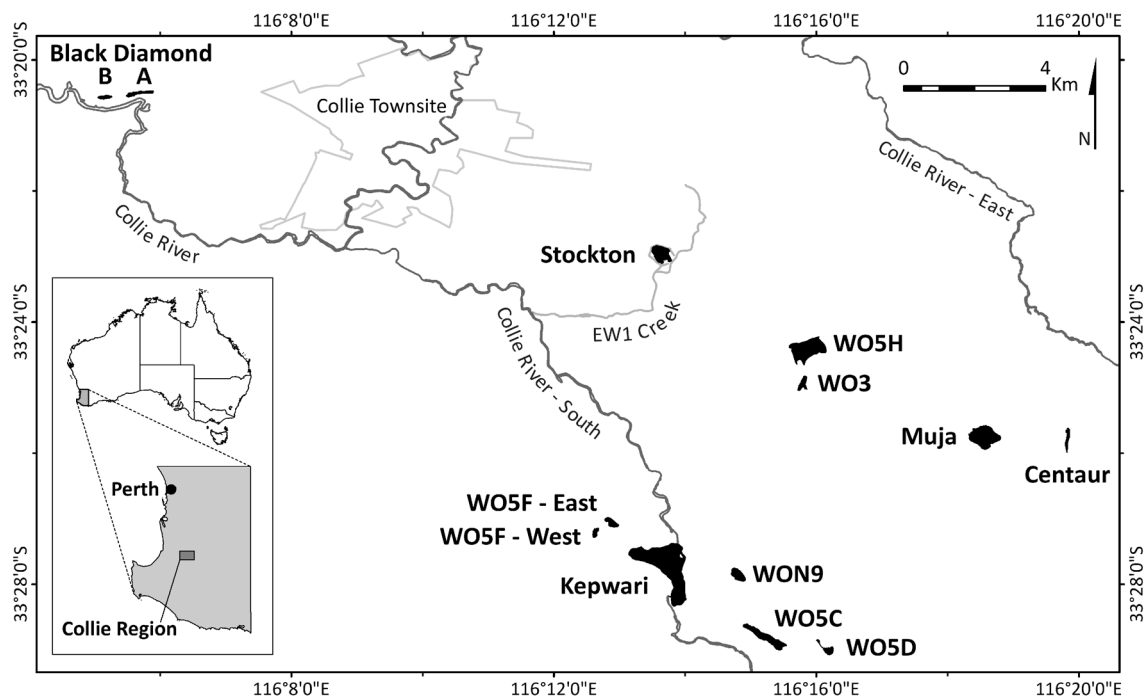
Fig. 1 Pit-lake district created by coal mining. Collie, Western, Australia. Study lakes were: WO5H, Kepwari and Stockton. Map by Mr. Karl Zwickl (Edith Cowan University). Created 2019

revegetated and the final intended use of the lake is as a recreational resource. The original closure plan was to keep the lake isolated from the Collie River South using the diversion; however, a large rain event in 2011 caused the river to breach the diversion and enter the lake, temporarily improving lake water quality by increasing the pH. The company was granted approval to connect the pit lake to the river permanently as part of the closure plan, which will see the lake relinquished back to the state of Western Australia (Lund and Blanchette 2018).

Field and Laboratory Protocols

Lakes WO5H, Stockton, and Kepwari were each sampled once in April 2013 (see "Data Analysis" section, below for experimental design). Macroinvertebrates and habitat features were collected from the littoral zone approximately every 200 m around the perimeter of each lake in an attempt to equalise sampling intensity among the lakes (Stockton $n = 8$, Kepwari $n = 21$, WO5H $n = 14$; perimeter distance determined using Google Earth © mapping tool). A surface water sample was collected from the centre of each lake for chemical analysis. Sediment for microbial and chemical analysis was collected from five sites at 2–3 m depth in each lake ≈ 5 m from the littoral zone (varying 1–2 m depending on prevailing conditions and safety for SCUBA divers). In situ measurements of surface physico-chemistry were collected at each site.

Macroinvertebrates were collected from a 0.5 m^2 quadrat via vigorous sweep sampling for 20 s using a square dip net with 500 μm mesh and immediately preserved in 70% ethanol. Macroinvertebrate identifications (to species, where possible) were completed according to Williams (1980), Davis and Christidis (1997), Hawking and Theischinger (1999), Suter (1999), Watts (2002), and Gooderham and Tsyrlin (2002). In this study, zooplankton captured in the quadrat were counted as macroinvertebrates as they can compose the bulk of pit lake biota (Gammons et al. 2009).

In situ measurements of physico-chemistry (turbidity (NTU), chlorophyll *a* (μg L^{-1}), pH, dissolved oxygen (DO; mg L^{-1}), % saturation, conductivity (EC; mS cm^{-1}), temperature (°C), and oxidation reduction potential (ORP; mV) were recorded using the Hydrolab DataSonde 4A (Hach Co., Loveland, CO, USA) ≈ 10 cm above the benthos (5–10 cm below the surface for macroinvertebrate transects and 2–3 m below the surface at sediment collection sites).

Water samples for metal and nutrient analysis were collected ≈ 10 cm below the surface of each lake. Each sample was split into three aliquots: one unfiltered and two filtered. Samples (where appropriate) were filtered through 0.5 μm Pall Metrigard™ filter paper under vacuum pressure. The unfiltered aliquot was frozen (− 12 °C) for later determination of total nitrogen (TN) and phosphorus (TP) following persulphate digestion as per APHA (1998a). One filtered aliquot was frozen (− 12 °C) for later determination of chloride (Cl$^-$) using ion chromatography (Methrohm, Switzerland),

Fig. 2 Mine-pit lakes. Collie, Western Australia, 2013. **a–c** Lake Stockton showing riparian trees (**a**), reeds (**b**), and mobile banks (**c**) typical of the lake. **d–f** Lake WO5H littoral planting in high (**d**) and low (**e**) densities, and broad lake aspect (**f**). **g–i** Lake Kepwari riparian (**g**, **h**) and across lake (**i**)

nitrate/nitrite (NO_x-N), filterable reactive phosphorus (FRP-P), and ammonia (NH_3-N) on a Lachat (Hach, USA) autoanalyser (APHA 1998b), and dissolved organic carbon (DOC; measured as non-purgeable organic carbon) using a total carbon analyser (Shimadzu, Japan). The second filtered aliquot was acidified with nitric acid (to a pH < 2 ≈ 1% v/v) and then kept at 4 °C for later determination by ICP-AES/MS of a range of metals/metalloids (Al, As, B, Ca, Cd, Cr, Co, Cu, Fe, Hg, K, Mg, Mn, Na, Ni, Pb, Se and Zn) and S (APHA 1998b).

A 20 m riparian transect was established parallel to each macroinvertebrate collection site to record percent bank cover of grass, leaf litter cover, twigs/branches, exposed soil, canopy cover, standing dead trees, rushes, and shrubs as well as the extent of undercutting, slumping, gullying, and bank slope. Benthic cover was also quantified, including periphyton/algal mat, aquatic and semi aquatic macrophytes, leaf litter, and coarse particulate organic matter (CPOM). A

Krumbein phi-scale was used to categorise sediment composition with local modifications for data analysis (sediment size index 1: sand-dominated, few rocks > 0.5 cm diameter (d), 2: silt-dominated, few rocks > 0.5 cm d, 3: sand and silt, few rocks > 0.5 cm d, 4: complex rocky mix—little sand/silt, rocks < 25 cm d, 5: boulders present > 25 cm d (Blanchette and Lund 2017; Blanchette and Pearson 2012, 2013).

Sediments were collected by SCUBA divers using modified gamma-sterile syringes, ensuring that only the top 5 cm was retained for microbial and chemical analysis (from Blanchette et al. 2019). Samples were immediately put on ice for transport, kept at − 20 °C for short-term field storage, and finally − 80 °C in the lab. Benthic sediments (7 to − 3 on phi scale) were analysed for bioavailable metals and nutrients according to U.S. EPA protocol 3050B (USEPA 1996) at the Edith Cowan University Analytical Chemical Laboratory using the Thermo Fisher Scientific iCAP QICPMS and 7600 ICPOES (Blanchette et al. 2019). Total N (Rayment

and Lyons Method 7A5) and total organic C (acidification method 6B1) (Rayment and Lyons 2011) were analysed at CSBP Perth, Australia.

Microbiome Analysis

DNA was extracted from 750 μg of wet sediment using the FavorPrep™ Soil DNA Isolation Mini Kit and quantified using a Qubit® 3.0 Fluorometer (Thermo Fisher Scientific Inc.) according to manufacturers' protocols. Microbiome analysis (16S rRNA sequencing) was performed by the Lotterywest State Biomedical Facility Genomics at the University of Western Australia according to Caporaso et al. (2012) with local modifications and detailed in Nagel et al. (2016).

Samples (1 ng) were amplified using the 16S V4/5 primers (515F: GTGCCAGCMGCCGCGGTAA and 806R: GGACTACHVGGGTWTCTAAT). To minimise primer–dimer formation and streamline downstream purification we used a mixture of gene-specific primers and gene-specific primers tagged with Ion Torrent-specific sequencing adaptors and barcodes (ratio of tagged and untagged primers was 90:10). Using this method, the ≈ 10-cycle inhibition observed by using long-tagged primers could be reversed and all samples were amplified using 18–20 cycles (5PRIME HotMasterMix, 5PRIME, USA) and confirmed by agarose gel electrophoresis. PCR product formation was quantified with a Qubit, as above. A maximum of 100 samples were diluted to equal concentrations and adjusted to a final concentration of 15 pM. Templated ion sphere particles were generated using an Ion Chef (Thermo Fisher Scientific) with a 400 bp templating kit and sequenced on an Ion PGM (Thermo Fisher Scientific) (850 cycles, 400 bp sequencing kit, modal read length of 309 bp). Reads were trimmed to improve quality using Torrent Suite 5.0 (Thermo Fisher Scientific). Taxonomic names were assigned to operational taxonomic units (OTUs) using software analysis programme Quantitative Insights into Microbial Ecology (QIIME, version 1.7) (Caporaso et al. 2010). OTUs based on 97% similar specific 16S rRNA sequence identities were used to distinguish different 'species' (or OTUs) of microbes using Greengenes database (McDonald et al. 2012; version 12_10). Samples were rarefied at 5000 reads for calculating dissimilarity between samples using UniFrac metrics.

Data Analysis

The overall experimental design incorporated collection of samples for aquatic macroinvertebrates and benthic microbes (Archaea, Bacteria) from three lakes during April 2013. To facilitate interpretation, biophysical and chemical data were separated into different matrices for analysis: (1) biotic (aquatic habitat and riparian characteristics), (2) water quality (physico-chemical and water chemistry), and (3) sediment chemistry (sensu Blanchette and Lund 2017). Biotic and in situ physico-chemical data were collected at each macroinvertebrate transect, and in situ physico-chemical data and sediment chemistry data at each microbe collection site. We tested the null hypotheses of no significant difference in water quality variables among lakes and no significant differences in macroinvertebrate or microbe assemblages among lakes. Biotic, water quality, sediment chemistry, and organism data were then ordinated to illustrate multivariate patterns and correlated against ordination axes to elucidate highly-correlated taxa. Correlations between biotic, water quality, and sediment chemistry variables and organism data were first tested for significance before relationships with specific parameters were explored.

Biotic, water quality, and sediment chemistry were compared among lakes using PERMANOVA in PRIMER-e with the Euclidean distance measure on normalised data (Anderson et al. 2008). Lake was treated as a fixed factor and transect was a random factor nested within lake (9999 permutations, significant $p < 0.05$), and pseudo-F is reported as F (Anderson et al. 2008). Biotic, water quality, and sediment chemistry data were ordinated with principal components analyses (PCA) in PRIMER-e. Data were predominantly linearly related to ordination axes and the significance of this relationship was assessed using Pearson's r ($\alpha = 0.01$) (Blanchette and Pearson 2012).

While 43 macroinvertebrate samples were collected from all three lakes (as above), two transects (one each from Stockton and WO5H) contained no animals and were thus excluded from analysis (41 samples in final analysis; Stockton $n = 7$, Kepwari $n = 21$, WO5H $n = 13$). Rare macroinvertebrate taxa ($< 0.01\%$) were excluded to decrease noise in the data set and improve detection of correlations between assemblages and biophysical variables (Blanchette and Pearson 2012; Cao et al. 2001; Herlihy et al. 2005; Marchant 2002), reducing an initial count of 40 taxa (across all lakes) to 19. The results therefore only describe common taxa. Macroinvertebrate species richness (S; common taxa only) was determined as mean S per quadrat per lake, and was calculated using one-way ANOVA in SigmaPlot 13.0 with a Tukey pairwise test (Systat Software, Inc.). Abundances were log (x)-transformed prior to analysis (Anderson et al. 2008; Blanchette and Pearson 2012). Macroinvertebrate log(abundance) data, hereafter referred to as 'assemblage data', were compared among lakes using PERMANOVA in PRIMER with the Bray–Curtis distance measure; again, lake was treated as a fixed factor and transect was a random factor nested within lake (9999 permutations, significant $p < 0.05$) and pseudo-F is reported as F (Anderson et al. 2008). Macroinvertebrate assemblage data were ordinated with nonmetric multidimensional scaling (NMDS) in PRIMER-e—a 2- or 3-dimensional solution was optimal

for all runs (stress below 0.20). Pearson–Kendall correlation (Pearson's r, as per biophysical variables, above) was used to identify relationships between macroinvertebrate assemblage data with NMDS axes (Blanchette and Pearson 2012; Herlihy et al. 2005). Correlations between macroinvertebrate assemblage data and biotic, water quality and sediment chemistry data were analysed using RELATE (Spearman's rank; r_s, $p < 0.05$; 9999 permutations) with a Pearson's correlation in PRIMER-e.

While 15 sediment samples were collected from all three lakes (as above), two transects (one each in Stockton and WO5H) did not contain sufficient DNA to amplify for downstream application and were thus excluded from analysis (13 samples in final analysis; Stockton $n = 4$, Kepwari $n = 5$, WO5H $n = 4$). Hypothesis tests on OTU (operational taxonomic unit) weighted (abundance and presence/absence) UniFrac genetic distance data were conducted with a PERMANOVA in PRIMER-e and ordinated with NMDS (factors, permutations and stress as per macroinvertebrate data). Correlations between OTU data and sediment chemistry and water quality were analysed using RELATE as per macroinvertebrate assemblage data and Blanchette et al. (2019).

Results

Biophysical Variables

Aquatic and riparian characteristics (biotic parameters; Supplementary Table 1) were significantly different across lakes (PERMANOVA; $F_{(2, 38)} = 3.96$, $p < 0.001$), and biotic parameters at all lakes were significantly different from each other (PERMANOVA pair-wise; all lake comparisons $p < 0.001$). A PCA plot (Axis 1; 18.6% of the variance, Axis 2; 16.2%, Axis 3; 12.8%) separated Lake Stockton from lakes Kepwari and WO5H along Axes 1 and 2 (Fig. 3). Pearson–Kendall correlation analysis ($|r| = _{0.01, 40} \geq 0.393$) indicated that PCA axis 1 was most strongly correlated with bank geomorphology (slumping, undercutting, slope; $|r| > 0.700$), PCA axis 2 was most strongly correlated with vegetation (rushes, riparian tree condition; $|r| > 0.670$), and PCA axis 3 was most strongly correlated with leaf litter ($|r| > 0.629$). Relative to Lakes Kepwari and WO5H, Stockton had potentially more bank instability, a greater cover of aquatic plants/rushes, and less terrestrial leaf litter input (Supplementary Table 1).

In situ water quality (Table 1) and water chemistry (Table 2) data were significantly different across lakes (PERMANOVA; $F_{(2, 38)} = 154.47$, $p < 0.001$), as were pairwise comparisons of these data between all lakes ($p < 0.001$). A
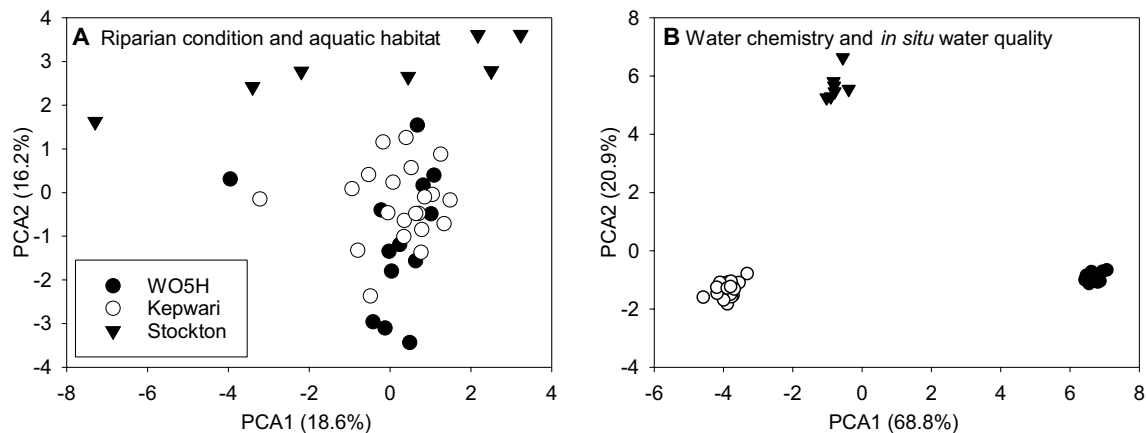

Fig. 3 Principal components analysis (PCA) ordinations of biophysical variables from pit lakes in Collie, Western Australia. Figure **a** shows results of an ordination for riparian condition and aquatic habitat and **b** shows results of a different ordination for water chemistry and in situ water quality

Table 1 In situ biophysical data from sites in Collie mine lakes, April, 2013

	Turbidity (NTU)	Chl. a (µg L^{-1})	pH	DO (mg L^{-1})	Sat. (%)	Cond. (mS cm^{-1})	Temp. (°C)	ORP (mV)
WO5H (n = 13)	0–2.6	0.33–1.31	2.66–2.89	8.33–8.85	94.1–96.9	1.56–1.58	21.2–21.7	461–468
Kepwari (n = 21)	0–1.1	0–0.56	3.90–3.95	7.17–8.97	81.9–102.2	3.04–3.90	20.8–22.1	355–386
Stockton (n = 7)	0–6.6	0.12–0.54	3.62–3.67	8.08–9.09	90.9–100.1	0.618–0.621	20.8–21.2	427–438

Sites correspond to macroinvertebrate collection transects

Table 2 Surface water chemistry data from Collie lakes collected April, 2013

	Al (mg L^{-1})	As (mg L^{-1})	B (mg L^{-1})	Ca (mg L^{-1})	Cd (mg L^{-1})	Co (mg L^{-1})	Cr (mg L^{-1})	Cu (mg L^{-1})	Fe (mg L^{-1})	Hg (mg L^{-1})	K (mg L^{-1})	Mg (mg L^{-1})	Mn (mg L^{-1})
WO5H	21.7	0.0061	0.0970	39.0	0.0024	0.1829	0.0086	0.0084	3.63	< 0.00005	2.3	40.3	1.7320
Kepwari	1.4	0.0008	0.1440	53.4	0.0004	0.0260	0.0005	0.0005	0.19	0.00009	1.3	88.7	0.1840
Stockton	2.6	0.0006	0.0590	10.7	0.0006	0.0354	0.0006	0.0019	0.17	0.00007	1.8	14.3	0.1470

	Na (mg L^{-1})	Ni (mg L^{-1})	Pb (mg L^{-1})	S (mg L^{-1})	Se (mg L^{-1})	Zn (mg L^{-1})	NH$_4^+$ (mg L^{-1})	NO$_x$ (mg L^{-1})	Total N (mg L^{-1})	FRP (mg L^{-1})	Total P (mg L^{-1})	Cl$^-$ (mg L^{-1})	DOC (mg L^{-1})
WO5H	116	0.2369	0.0080	145.9	0.0049	2.3510	0.4980	0.0580	1.06	< 0.002	< 0.02	190	0.9
Kepwari	397	0.0351	0.0039	35.1	0.0029	0.5470	0.0060	0.5800	0.827	< 0.002	< 0.02	898	1.1
Stockton	77	0.0516	0.0014	16.6	0.0011	0.4730	< 0.0030	0.2510	0.515	< 0.002	< 0.02	111	1.0

	Total anions (mEq)	Total cations (mEq)
WO5H	14.483	13.140
Kepwari	27.572	27.452
Stockton	6.275	5.443

N 1 per lake, *NO$_x$* NO$_2$/NO$_3$, *FRP* filterable reactive P

PCA plot (Axis 1; 69.9% of the variance, Axis 2; 19.6%, Axis 3; 4.8%) separated all lakes along axis 1, whereas Lake Stockton was separated from lakes Kepwari and WO5H along axis 2 (Fig. 3). Pearson–Kendall correlation analysis ($|r| =_{0.01, 40} \geq 0.393$) indicated that PCA axis 1 was most strongly correlated with metal concentration, sulphur, ammonium, and pH [e.g. Ni, Al, Fe, Mn, Zn, pH, sulphur, $NH_4^+ |r| > 0.940$) and axis 2 was correlated with Ca ($|r| = 0.940$)]. Lake WO5H had higher concentrations of metals, sulphur, and ammonium, and a lower pH than lakes Stockton and Kepwari. Lake Stockton had the lowest Ca concentration (Table 1, Supplementary Table 2).

Sediment chemistry data were not significantly different across lakes (PERMANOVA; $F_{(2, 10)} = 1.96$, p = 0.066) (Table 3).

Macroinvertebrates

In total, 81,466 individuals from 19 aquatic taxa were identified across all lakes (Supplementary Table 2). Planktivores composed 98.5% of the individuals collected, which were mainly calanoid copepods (98.4%) from lakes Kepwari and Stockton. Calanoid copepods were not collected from WO5H. The detriti-herbivore Chironomini (Chironominae) was the most abundant insect taxon (1.80%), with high abundances in Lake WO5H and few individuals found in lakes Kepwari and Stockton. Predators were rare (1.28%), but also were the most diverse group, even at high taxonomic levels within the Insecta (Diptera, Odonata, Coleoptera) as well as the Arachnida (Acarina; Orabatidae). Species richness (*S*) was significantly different among lakes ($F_{(2,40)} = 8.76$, p < 0.001), and results of a Tukey test indicated that mean species richness per quadrat between lakes Kepwari and Stockton (p < 0.001) and WO5H and Stockton (p = 0.034) were significantly different ($S \pm SE$: Kepwari; 7.29 ± 0.55, Stockton; 3.25 ± 0.77, W05H; 5.93 ± 0.56).

Overall, macroinvertebrate assemblages were significantly different among lakes (PERMANOVA; $F_{(2, 38)} = 26.35$, p < 0.001), and assemblages from all lakes were significantly different from each other (PERMANOVA pairwise; all lakes p < 0.001). An NMDS ordination of lake macroinvertebrates (2-dimensional solution, stress = 0.09) showed strong separation of Lake WO5H from lakes Kepwari and Stockton (Fig. 4). Pearson–Kendall correlation analysis ($|r| =_{0.01, 40} \geq 0.393$) indicated that, although relatively rare, predators and omnivores tended to have high r-values relative to NMDS axes and were associated with Lake WO5H (NMDS axis 1; *Orthetrum caledonicum* (Libellulidae), Chironomini (Chironominae), *Necterosoma darwini* (Dytiscidae), *Ecnomina* sp. (Trichoptera), $|r| > 0.500$, NMDS axis 2; Ceratopogoninae (Ceratopogonidae), $|r| = 0.547$).

Results of a RELATE procedure for macroinvertebrate assemblages and aquatic habitat and riparian characteristics

Table 3 Sediment chemistry data from sites in Collie mine lakes, April, 2013. Sites correspond to microbe collection points

|	Total N (%)	Total C (%)	P (µg g⁻¹)	Na (µg g⁻¹)	Mg (µg g⁻¹)	K (µg g⁻¹)	Ca (µg g⁻¹)	Al (µg g⁻¹)	Cd (µg g⁻¹)	Co (µg g⁻¹)	Cr (µg g⁻¹)	Cu (µg g⁻¹)	Fe (µg g⁻¹)	Mn (µg g⁻¹)	Ni (µg g⁻¹)	Pb (µg g⁻¹)	S (µg g⁻¹)	Zn (µg g⁻¹)
K1	0.04	0.79	60.5	139.1	139.0	275.6	130.3	28051.5	40.0	9.4	0.55	5.7	2198.6	8.8	3.51	0.03	552.0	2.8
K2	0.12	3.74	129.1	4840.4	1541.6	1100.5	677.1	114524.9	360	44.1	3.06	35.3	44874.8	36.0	22.20	0.17	3061.4	11.8
K3	0.15	2.31	62.4	621.2	364.3	537.7	207.4	47438.9	43.6	16.9	1.07	8.8	8120.9	19.8	4.66	0.03	1068.8	3.6
K4	0.03	0.31	20.2	296.8	121.8	59.8	69.1	9517.1	12.1	6.2	0.16	2.4	4971.2	7.1	1.30	0.03	161.8	1.3
K5	0.04	0.61	55.6	427.2	365.8	251.9	167.9	41586.3	14.5	21.7	0.17	6.5	6594.5	11.8	2.59	0.03	176.2	4.8
S1	0.05	0.41	75.5	77.2	131.6	311.0	54.6	69554.0	15.8	25.7	0.21	12.8	3125.5	8.7	3.23	0.03	210.9	9.9
S3	0.13	2.28	132.8	153.4	270.1	515.3	124.2	105995.9	49.7	57.9	0.92	24.1	5000.8	12.1	6.11	0.08	920.1	13.0
S4	0.07	2.12	131.1	67.3	201.4	404.6	194.4	20106.0	23.2	16.8	0.22	7.7	1718.3	20.3	2.50	0.03	219.9	10.8
S5	0.07	14.50	106.9	83.4	153.7	414.5	172.8	19857.0	27.2	28.5	0.71	12.0	8036.1	12.0	3.60	0.11	706.2	28.4
WO5H1	0.02	0.20	25.9	56.1	39.3	94.4	77.3	12154.2	15.3	6.8	0.38	1.6	8804.2	7.3	0.53	0.03	378.0	4.7
WO5H2	0.03	0.13	33.8	62.1	46.9	118.3	55.5	18416.4	16.9	11.6	0.46	2.5	11751.2	15.2	0.68	0.03	455.4	6.0
WO5H3	0.04	0.26	15.8	37.7	19.0	23.3	98.0	1655.5	14.0	1.4	0.10	0.4	1193.9	1.9	0.16	0.03	97.9	1.2
WO5H4	0.03	0.15	5.0	27.2	24.8	11.9	66.2	2025.2	10.4	1.4	0.07	0.7	1055.9	0.0	0.21	0.03	65.9	1.5
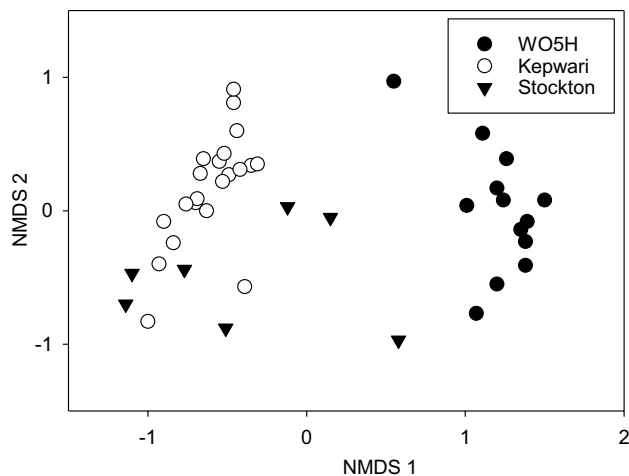
Fig. 4 Nonmetric multidimensional scaling (NMDS) ordination of macroinvertebrate assemblages from pit lakes in Collie, Western Australia collected April 2013. 2-D configuration, stress = 0.09. Assemblages were significantly different among lakes ($F_{(2, 38)} = 26.35$, $p < 0.001$)

Fig. 5 Nonmetric multidimensional scaling (NMDS) ordination of microbe assemblages (Archaea, Bacteria) from pit lakes in Collie, Western Australia collected April, 2013. 2-D configuration, stress = 0.11. Assemblages were significantly different among lakes ($F_{(2, 10)} = 2.38$, $p < 0.001$)

indicated that the relationship was significant ($r_s = 0.160$, $p = 0.018$). Macroinvertebrate assemblages on NMDS axis 1 were correlated with CPOM and benthic leaf litter cover ($|r| > 0.500$) in Lake WO5H, and aquatic plants ($|r| = 0.394$, NMDS axis 2) in Lake Stockton. There was also a significant relationship between macroinvertebrate assemblages and water quality (RELATE; $r_s = 0.728$, $p < 0.01$). On NMDS axis 1, assemblages in Lake WO5H were correlated with higher concentrations of metals (Cd, Ni, Co, Al, Cu, Cr, Fe, $|r| > 0.931$) and lower pH ($|r| = 0.932$), whereas on NMDS axis 2 assemblages in Lake Stockton were correlated with lower Ca concentrations and lower conductivity measurements ($|r| > 0.460$).

Microbes

Microbes (UniFrac distance) were significantly different among lakes (PERMANOVA; $F_{(2, 10)} = 2.38$, $p < 0.001$), and assemblages from all lakes were significantly different from each other (all pairwise comparisons were $p < 0.05$). Similar to the lake macroinvertebrates, an NMDS ordination of lake microbes (2-dimensional solution, stress = 0.11) showed separation of Lake WO5H from lakes Kepwari and Stockton (Fig. 5).

Across all lakes, there were 298 unique OTU categories from 178 families of microbes, seven of which were from kingdom Archaea (most were bacteria). Most microbes were rare; 281 of the 298 OTUs had average relative abundances (RAs) of less than 0.01, 17 OTUs had an average RA of 0.01–0.06, and one OTU had an average RA of 0.30, which was an unidentified bacterium (Fig. 6). Of the top nine OTUs by RA (Fig. 6), only seven of the taxa could be identified
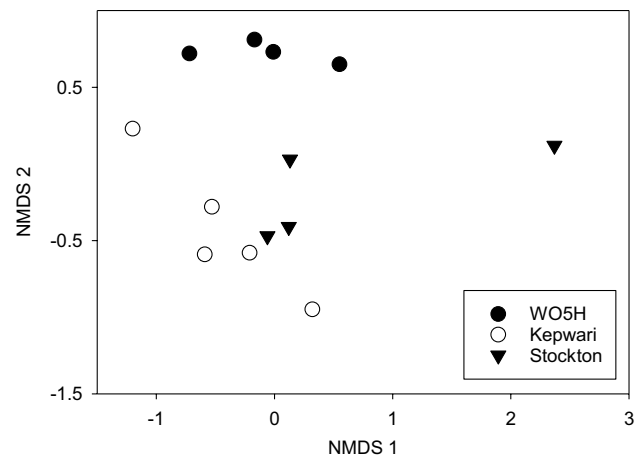
to class and one could be identified to family. OTU (species) richness (S) was significantly different among lakes ($F_{(2, 10)} = 5.40$, $p = 0.026$), which was driven by a significant difference in mean S per sample between lakes Kepwari and WO5H (Tukey's test; $p = 0.024$, $S \pm$ SE: Kepwari; 139.4 ± 23.9, Stockton; 112.8 ± 18.0, W05H; 100.0 ± 7.39).

Results of a RELATE procedure for microbes (UniFrac distance) and aquatic chemistry (in situ and analyses) indicated that the relationship was significant ($r_s = 0.456$, $p < 0.001$). Similar to the macroinvertebrates, microbes in Lake WO5H were correlated (Pearson's r; $|r| =_{0.01, 13} \geq 0.641$) with higher metal concentrations (NMDS axis 2; Cu, Cd, Ni, Co, Al, $|r| > 0.860$) and lower pH ($|r| = 0.870$). Unlike the macroinvertebrates, Ca and in situ conductivity were not significantly correlated with NMDS axes. Instead, turbidity was the only significantly correlated water quality variable associated with NMDS axis 1 ($|r| = 0.840$), separating Lake Stockton from lakes Kepwari and WO5H (n.b., although partially driven by a highly turbid data point in Lake Stockton). The relationship between microbes (UniFrac distance) and sediment chemistry was not significant (RELATE; $r_s = -0.016$, $p = 0.47$).

Discussion

The broad aim of this research was to compare the environmental drivers of a routinely-used bio-assessment tool (macroinvertebrates) with those of microbial assemblages (Archaea, Bacteria) in acidic pit lakes. The purpose of this comparison was to determine if microbes, which are rarely used in freshwater bio-assessment programs, provide

Fig. 6 Top nine operational taxonomic units (Archaea, Bacteria) by relative abundance from pit lakes in Collie, Western Australia collected April, 2013
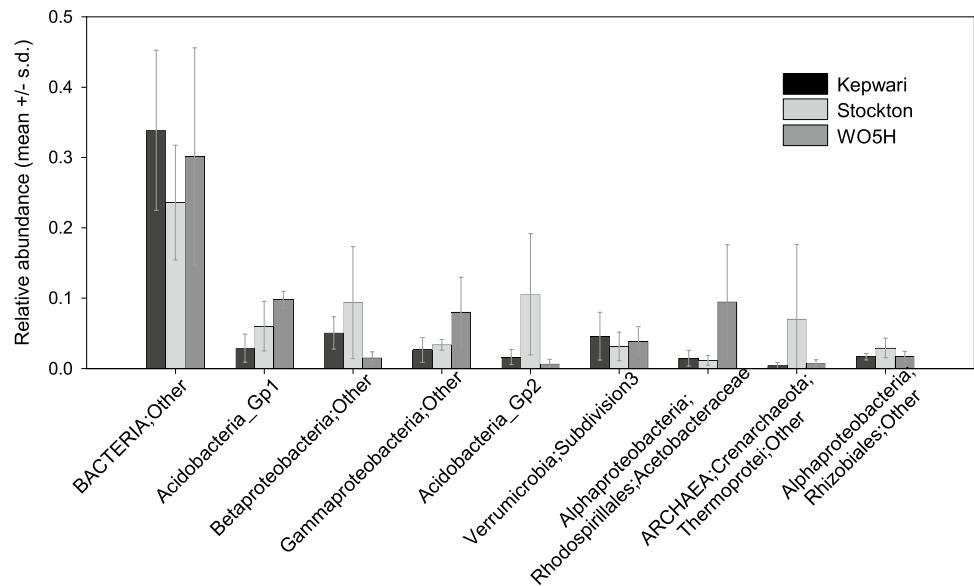
complementary or alternative understanding of pit lake ecosystems. We found that in many ways, the two groups were complementary; their assemblages were significantly different across lakes and correlated with similar water quality and biotic characteristics, particularly the metals in Lake WO5H. This is an encouraging step towards developing a standardised collection protocol for benthic microbes—critical for progressing any new monitoring program. However, we were unable to identify many of the microbes, and in this respect, the macroinvertebrates provided more information regarding ecosystem processes. Given the extensive body of literature regarding the identification and ecology of macroinvertebrates, this research indicated that aquatic macroinvertebrates would be more useful in pit lake monitoring programs than microbes. Nevertheless, rapid advances in technology and decreases in per sample costs suggest that microbes as environmental bio-indicators is a line of enquiry worth pursuing.

Biophysical variables were significantly different across lakes, which reflected variable riparian conditions and groundwater characteristics. Lakes Kepwari and WO5H had more input of terrestrial leaf litter and more gently sloping banks than Lake Stockton, where signs of bank instability were more evident (i.e. slumping, undercutting, gullying). Bank instability in Lake Stockton may have been partially due to erosive wave action from recreational water skiing. Relative to other lakes, Stockton had a greater cover of true aquatic plants, but cover of both erosion and plants was patchy, and plants occurred in areas of limited erosion. Metal concentrations (e.g. Al, Cd, Cr, Ni, Zn) in Lake WO5H were high, likely because most of the drainage from the adjacent mine site collects in the lake were from surface run-off or interflows. Aluminium concentrations were particularly high at 21.7 mg L^{-1}, although Cd, Cr, Ni and Zn exceeded the

ANZECC/ARMCANZ (2000) guideline values for the 95% protection of aquatic ecosystems by up to 300%.

Macroinvertebrate assemblage composition and species richness was significantly different across all lakes, despite the close proximity of lakes within the 'lake district' and the prevalence of hardy, cosmopolitan taxa in the region (Smith et al. 1999). Across all lakes, predators were rare, but diverse, and calanoid copepods were the most relatively abundant organism collected (at 98.4%), but were not found in Lake WO5H. Planktonic communities may dominate pit lake biota, and their prevalence depends heavily on water quality (see Gammons et al. 2009). Lake WO5H was distinct from the other two study lakes by having high metal concentrations, a well-known inhibitor of zooplankton establishment in mine-affected oligotrophic lakes (Roch et al. 1985). Even relatively low metal concentrations in acidic and/or saline mine water can be deleterious to zooplankton, either directly or by compromising phytoplankton food items (Leppänen et al. 2017). While this was not a food web study per se, the total absence of calanoid copepods (the most dominant taxon of the study) at Lake WO5H indicates that the invertebrate-planktonic food web in WO5H was different from those of Lakes Kepwari and Stockton.

Relative to the other study lakes, Lake WO5H contained more predators and detriti-herbivores, which were significantly correlated with CPOM and benthic leaf litter cover. Lake WO5H also had a mean species richness nearly double that of Lake Stockton (but less than Lake Kepwari). These results suggest that despite poor water quality indicators of low pH and high metal concentrations, nutrient input from terrestrial leaf litter can support or augment pit lake ecosystems (sensu Blanchette and Lund 2016). After post-abandonment pit contouring, Lake WO5H underwent a planting program where native terrestrial seed was introduced

to the pit below the final water level, allowing plants to be flooded as the pit filled (see Lund et al. 2014). Input of terrestrial leaf litter—as habitat and food—is a critical function of riparian zones, and riparian structure appears to be the main factor influencing leaf litter input (Naiman and Decamps 1997). Mine pit lake catchments have traditionally been considered risks due to acid mine drainage, public safety, and discharge (McCullough and Lund 2006). This research indicates that pit lake riparian zones can also provide an opportunity to enhance aquatic biodiversity, despite potentially poor water quality. While this study didn't test if organisms were responding to terrestrial leaf litter as either 'habitat' or 'food' (e.g., Karádi-Kovács et al. 2015; Richardson 1992), future research will employ techniques such as stable isotope analysis to determine basal food web sources and relationships among taxa.

Despite differences in aquatic chemistry, measured sediment chemistry was not different among lakes. Compared to water quality variables, sediment chemistry was extremely spatially patchy within lakes and our small sample size (necessitated by cost) may not have fully captured within-lake variability. In an acidic Ontario pit lake, metal accumulations in the sediments were also highly spatially variable, and likely due to localised discharges of groundwater upwelling—discharge locations were 'encrusted' with iron and co-precipitated metals occurred directly next to sediments with low metal concentrations (Kalin and Wheeler 2013). Lake sediments are considered a "record of lake history" (Jones and Bowser 1978), and even in these apparently simple systems, depositional and hydrological processes produce a complicated benthic system. Future work will attempt to elucidate how variables such as sedimentation and mineralisation affect the within-lake spatial variability of sediments.

We took a classic ecological approach to the microbial study: collect organisms and biophysical variables, then statistically analyse and determine any correlations in the data. This approach worked well in seasonal rivers in New South Wales (Blanchette et al. 2019). However, in pit lakes sediment, microbial assemblages were not correlated with sediment chemistry, but rather water quality. One possibility is that we did not target the variables that actually drove differences in pit lake microbial assemblages—such as the precise nutrient fraction. Another possibility is that the microbes we collected were predominantly from the pelagic fraction of the food web, rather than the sediments. The benthos of the study lakes were mainly composed of kaolinite clay, which are powdery and mobile. It would be difficult for a distinct benthic biofilm community to form on a mobile sediment with little integrity (Gerbersdorf and Wieprecht 2015). Interstitial pore water and handling error by divers could have also introduced pelagic DNA to samples. Essentially, if the bottom of a lake is considered a 'depositional reservoir'

(sensu Jones and Bowser 1978), we may have sequenced the DNA of dead/dying and/or transient pelagic microbes, rather than a true benthic community. Future work will collect pelagic microbes at different depths as well as 'benthic' microbes (as in Blanchette et al. 2019) and identify assemblages from sediment cores at different depths.

As discussed above, microbial assemblages in pit lakes were correlated with water chemistry, particularly metals and low pH, which separated Lake WO5H (which had the lowest OTU richness) from lakes Kepwari and Stockton. Microbial assemblages were also correlated with high turbidity in Lake Stockton. Many taxa were unable to be identified beyond class, and the most abundant taxon was an unidentified bacterium. However, in a September 2014 study of microbial assemblages in the Collie River South Branch (which flows through Lake Kepwari—see Lund et al. 2018), using the methods above, we (the authors) described 1239 OTUs from 591 different microbial families (Bacteria and Archaea; 428 ± 126 OTU richness per sample), most of which could be identified to genus (Lund and Blanchette 2018). In the river, bacteria unidentified at the phylum level were extremely rare in samples. Data from the river demonstrate that DNA techniques and libraries were sufficient to sequence and identify benthic aquatic microbes in the region.

Relative to co-occurring natural waterbodies, microbial assemblages in these acidic pit lakes were depauperate, and many did not occur in DNA databases at the time of analysis. While it is challenging to compare OTU richness among studies due to the potential effects of methodology, acidic pit lakes in Collie have similar OTU richness (100–130 per sample) to environments such as sea ice (128 OTU richness per sample; Yergeau et al. 2017), and hot springs (100–300 per-sample OTU richness; Colman et al. 2016). Similar to this study, a large proportion (up to 20%) of bacteria from a Chilean desert salt lake could not be identified to phylum level (Rasuk et al. 2016), indicating that in extreme environments, it is not unusual to find large proportions of unclassified bacteria in samples.

Conclusions

As acidic freshwater systems with high metal concentrations and relatively depauperate microbe communities, the Collie pit lakes could be considered extreme environments. However, as the macroinvertebrate data shows, the addition of even small amounts of carbon in the form of terrestrial nutrients altered biodiversity. In Lake WO5H, trees planted below the eventual water line added nutrients in the form of terrestrial leaf litter to the aquatic ecosystem. This can have profound implications for remediation, where connection to catchments can be a fundamental part of mine

closure planning (Blanchette and Lund 2016). Drawing on the significant body of wetland restoration literature, sculpting the sides of pit lakes to form littoral zones for semi-aquatic plants can provide nutrients and habitat, but can also function as the 'kidneys' of lakes with poor water quality, filtering undesirable elements and stabilising banks (Mitsch 2012; Mitsch et al. 1998, 2012, 2015).

Macroinvertebrate and microbial assemblages differed across lakes and correlated with some similar water quality and biotic characteristics, particularly the metals in Lake WO5H. In this respect, the information provided by the macroinvertebrates and microbes was complementary. Ecological indicators are only useful if there are clear links between the taxon and ecological role (Kurtz et al. 2001). As many of the pit lake sediment microbes were unidentified, we could not even begin to speculate on ecological function, even of the most abundant taxon in the study. Nevertheless, technology is advancing quickly and microbes are attractive bio-indicators due to rapid sample collection and analysis. Future environmental impact programs are likely to include microbes as they become increasingly attractive assessment tools (Lau et al. 2015).

Compliance with Ethical Standards

Ethical approval This research was conducted according to the Australian Code for the Responsible Conduct of Research and the Edith Cowan University Framework for the Responsible Conduct of Research.

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